Complete Genome Sequences of *Cellvibrio japonicus* Strains with Improved Growth When Using α-Diglucosides

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**ABSTRACT** *Cellvibrio japonicus* is a saprophytic bacterium that has been studied for its substantial carbohydrate degradation capability. We announce the genome sequences of three strains with improved growth characteristics when utilizing α-diglucosides. These data provide additional insight into the metabolic flexibility of a biotechnologically relevant bacterium.

*Cellvibrio japonicus* Ueda107 (NCIMB 10462) is a Gram-negative bacterium that was isolated from Japanese soil in 1952 (1–3). *C. japonicus* possesses over 200 genes that encode carbohydrate-active enzymes (CAZymes), many of which have been studied for applications in renewable energy (4–6). While the bacterium exhibits a robust carbohydrate utilization metabolism, the Ueda107 strain exhibits an extended lag phase when using α-diglucosides (Fig. 1). To select for *C. japonicus* strains with improved growth when using kojibiose, nigerose, or isomaltose as a sole carbon source, we grew the wild-type bacterium for 48 h in a single disaccharide (0.5% [wt/vol]) using microplate growth parameters identical to those previously published (7, 8). After 48 h, the cultures were reinoculated (1:100 dilution) back into the same carbon source, which resulted in growth phenotypes similar to those observed using glucose (Fig. 1). After 48 h of growth, 100 µl of each of the three reinoculated strains was individually recovered, mixed with 100 µl of 50% sterile glycerol, and frozen at −80°C for long-term storage.

To obtain sufficient cell mass for DNA extraction, we used the protocols published for *C. japonicus* Ueda107 RNA sequencing (RNA-seq) analysis (9–12). Genomic DNA was then extracted using a QiAamp high-throughput (HT) DNA kit (Qiagen, Hilden, Germany) and fragmented with an LE220 ultrasonicator (Covaris, Inc., Woburn, MA), which had an insert size between 253 and 267 bp. Genomic libraries were constructed using a Nextera XT library preparation kit (Illumina, Inc., San Diego, CA), following the manufacturer's instructions. DNA quality and quantity were determined using either Qubit fluorometric quantification (Thermo, Waltham, MA) or TapeStation DNA electrophoresis (Agilent, Santa Clara, CA) when appropriate, with the merits of each previously described (13). A MiSeq platform with 2 × 150-bp paired-end read sequencing was used to generate reads for each strain (Illumina, Inc.). Raw sequence data (.bcl files) generated from the Illumina MiSeq platform were converted into FASTQ files and demultiplexed using the Illumina bcl2fastq 2.17 software. Default software parameters were used, with the exception that one mismatch was allowed for index sequence identification. Adapter sequences were trimmed during this process, which resulted in an average read length of 125 bp. The mean Q score for all three strains was approximately 36%, and the processed reads were mapped and assembled using CLC Genomics Workbench v10.0, with the default software settings (Qiagen), against the published *C. japonicus* Ueda107 reference genome (GenBank accession number NC_010995). In all cases, 100% of the *C. japonicus* reference genome (4,576,573 bp) was covered, and the average G+C content of all three strains was 52%. The strain isolated from kojibiose medium (C. japonicus ADPT1-KOJIBIOSE) had 3,379,938 mapped reads (107-fold cover-
age), with a complete genome size of 4,576,591 bp (3,687 total genes). The strain isolated from nigerose medium (C. japonicus ADPT2-NIGEROSE) had 3,281,977 mapped reads (104-fold coverage), with a complete genome size of 4,576,586 bp (3,688 total genes). The strain isolated from isomaltose medium (C. japonicus ADPT3-ISOMALTOSE) had 3,204,888 mapped reads (101-fold coverage), with a complete genome size of 4,576,591 bp (3,687 total genes). All three strains were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (14). Given the similarity between the strains, a full comparative analysis may provide greater insight into the mechanisms for using α-diglucosides.

Data availability. The unique BioProject identifier for the entire study is PRJNA561085. The genome sequences were deposited in NCBI GenBank under accession numbers CP043304, CP043305, and CP043306. The raw data were deposited in the NCBI SRA under experiment numbers SRX6736265, SRX6736266, and SRX6736267.

ACKNOWLEDGMENTS

This work was supported by the U.S. Department of Energy, Office of Science, Office of Biological and Environmental Research, under award DE-SC0014183. The processing of C. japonicus cell pellets, library generation, genome sequencing, and quality control (QC) analysis was done by the company Genewiz (Plainfield, NJ) on a fee-for-service basis.

C.A.G. generated cell pellets for whole-genome sequencing and contributed to writing the manuscript. J.A.N. isolated the strains and contributed to writing the manuscript. J.G.G. supervised all aspects of the work and contributed to writing the manuscript. All authors read and approved the final submitted version of the manuscript.
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REFERENCES


